

THE QUEENSLAND MYCOLOGIST



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The Queensland Mycological Society

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Society Objectives

The objectives of the Queensland Mycological Society are to:

1. Provide a forum and a network for amateur and professional mycologists to share their common interest in macro-fungi
2. Stimulate and support the study and research of Queensland macro-fungi through the collection, storage, analysis and dissemination of information about fungi through workshops and fungal forays
3. Promote, at both the state and federal levels, the identification of Queensland's macrofungal biodiversity through documentation and publication of its macro-fungi
4. Promote an understanding and appreciation of the roles macro-fungal biodiversity plays in the health of Queensland ecosystems
5. Promote the conservation of indigenous macro-fungi and their relevant ecosystems.

Membership

Membership of QMS is \$25 per annum, due at the beginning of each calendar year, and is open to anyone with an interest in Queensland fungi. Membership is **not** restricted to people living in Queensland. Membership forms are available on the website, <http://qldfungi.org.au/>.

Please notify the membership secretary (memsec@qldfungi.org.au) of changes to contact details, especially your email address.

The Queensland Mycologist

The Queensland Mycologist is issued quarterly, **but issues may be combined if there is insufficient material for four**. Members are invited to submit short articles or photos to the editor for publication. It is important to note that it is a newsletter and not a peer-reviewed journal. However we do aspire to high standards of accuracy and there is an extensive review process.

Material can be in any word processor format, **but not PDF**. The deadline for contributions for the next issue is **31 October 2022**, but if you have something ready, please send it **NOW!** Late submissions may be held over to the next edition, depending on space, the amount of editing required, and how much time the editor has, or the newsletter may come out late.

Photos should be **submitted separately at full-size** to allow flexibility in resizing and cropping to fit the space available while minimising loss of quality. Authors who have specific preferences regarding placement of photos should indicate in the text where they want them, bearing in mind that space and formatting limitations may mean that it is not always possible to comply. Material from published sources (including internet sites such as Wikipedia) may be included **if that complies with copyright laws and the author and source are properly acknowledged**. However extensive verbatim copying is not acceptable.

Cover Illustration

Mutinus sp. (possibly *Mutinus bambusinus*) photographed by Ray Palmer in the Cairns area in January 2021. This unusual bifurcated specimen may have suffered damage during development. Photo © Ray Palmer.

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QMS activities

Meetings

Meetings are held on the second TUESDAY of the month at the National Heavy Vehicle Regulator, Level 3/76 Skyring Terrace, Newstead QLD 4006, from 6:30 – 8:30 pm. If you are parking underground at Gasworks Plaza, please remember that parking is free for two hours, entrance to the venue is on the ground floor of the Plaza, directly opposite Woolworths.

For those unable to attend, meetings can also be attended virtually on Zoom. Some meetings will be only on Zoom. Details will be provided via email.

Meetings are held monthly from February (no January meeting), unless otherwise scheduled. **Check the website for details and any changes.** There are typically 3-4 guest speakers invited during the year, with the other meetings informal. Suggestions from members for topics or names of potential speakers will be welcome at any time. Please contact a member of the Committee.

We like to publish notes from presentations in the *Queensland Mycologist*. However, the notes never do justice to the topic as they do not reflect the enthusiasm of the speaker or cover the discussion that follows, and not all talks are written up for the newsletter. If you do present at a meeting, a summary of that presentation that can be turned into a newsletter article will be greatly appreciated. But it is better to attend the meetings, get the information first hand, and participate in these invaluable information sharing opportunities.

Forays

QMS holds regular forays during the first half of the year. The dates are nominally the 4th Saturday of the month, but actual dates may vary and additional forays may also be held. Field trip details may change as a result of drought or other unforeseen circumstances. Check the website for changes.

Members are invited to suggest venues for additional forays. If you have any suggestions, (and especially if you are willing to lead a foray), please contact Wayne Boatwright or another member of the Committee.

Workshops

What do you, our members, want to learn more about that could be presented in a workshop? QMS is always on the lookout for workshop ideas. Members are encouraged to suggest topics, whether new or reruns of past workshops. Send your ideas to Wayne Boatwright (info@qldfungi.org.au).

Details of workshops will be included in newsletters and on the QMS website as they become available.

The Australasian Mycological Society

A reminder that the Australasian Mycological Society is well worth joining.

Their home page is at: <https://www.australasianmycologicalsociety.com/>

They also have virtual seminars. For more information visit the website.

QMS Program 2022

MONTH	MEETINGS	FORAYS/WORKSHOPS
Check the website for updates on QMS meetings and forays, and on the impacts of COVID-19 on QMS activities., and look out for emails from Wayne Boatwright.		
https://qldfungi.org.au/2022-calendar-year-monthly-meeting-space-dates-and-forays		
August	9 th : Wayne Boatwright – Forays of 2022	
September	13 th : Rod Rogers – Introduction to Lichens	
October	11 th : Dr Teresa Lebel – Truffles	
November	8 th : Dr Sally Fryar – Marine and Freshwater Fungi	
December	13 th : Christmas party. Details TBA	

Editor's Comments

Once again, a double issue. There will probably only be two issues of the newsletter this year, depending on my time and the amount of material I receive. I do have some material for the next edition, but need more. Hopefully I will be able to devote more time to the newsletter after I retire around the end of this year.

Ray Palmer sent some beautiful photos from the north late last year, and I decided they were worth pulling together for the newsletter. The cover features one of his photos.

Pat Leonard has prepared a Queensland Fungal Record of *Laccocephalum hartmannii*, as well as an article of the difficulties and pitfalls of trying to identify nondescript mushrooms, in this case “smelly brown mushrooms”. That task is not for the faint-hearted, and the article is a good guide to the sorts of actions required (and difficulties involved) as you work through the process of trying to identify fungi.

Fran Guard has prepared an article on the remarkable litter traps produced by *Crinipellis*. I would never have recognised them for what they are, so thanks Fran.

Invasive fungi are given too little attention unless, as with myrtle rust, they cause obvious harm to native

species or crops.

One very visible weedy fungus is the orange ping pong bat, *Favolaschia calocera*. There was an article on that species in the March 2009 newsletter, but it has spread significantly since then, so Theresa Bint's update on page 13 is most welcome. It is clear that there are still unanswered questions about the taxonomic status of the fungus we have in Australia, and in spite of the widespread and likely justified view that it is harmful, its actual ecological impact is not properly understood. Too few resources are allocated to the kinds of studies that are needed.

Irene Denton recently wrote *The Basics of Agaric Gills* for Barry Muir's excellent Fungi Foragers newsletter. The article is too big for us, and more importantly, I could never hope to recreate the excellent formatting, so I have suggested it be put on the QMS website.

Barry is based in Cairns so his newsletter focuses on that area. He emails it to any who want it, and you can ask to be added to Barry's mailing list via: unit57.may@gmail.com

Thank you to all who have contributed, both writers and reviewers. Enjoy!

New Species of Interest

Descriptions of three new species of *Agaricus* from Australia have been published in *Fungal Planet Descriptions*, part of the journal *Persoonia*. They are numbers 1400-1402.

Go to: <https://doi.org/10.3767/persoonia.2022.48.08>

The set of descriptions also includes three *Russula* species from Thailand (Nos 1423-1425), and *Serendipita petricolae* described by John Dearnaley et al (1426). To add to that, five pathogens of ratstail and related grasses (1427-1431) are described by the team I work with, led by Tracey Steinrucken (who is also current president of the Australasian Mycological Society) and Joe Vitelli. Most of the many fungi we have isolated from grasses (mostly from the genus *Sporobolus*) are new, illustrating just how much is still to be discovered. My role has been isolating and culturing the fungi, and trying to get them to sporulate (with mixed success!).

Laccocephalum hartmannii



Laccocephalum hartmannii © Pat Leonard (L), Warwick Nash (R)

Cap: planoconvex, circular in outline, occasionally reniform; 120 – 200 mm diameter; finely velutinous; brownish orange (7C7)¹ to burnt orange; margin acute and somewhat appendiculate.

Stipe: clavate; caespitose; central or eccentric (in the one population); 70 – 80 × 30 – 40 mm; velutinous; brownish orange.

Pores: decurrent; white; angular, 3 – 5 per mm; bruising brown; drying pale brown; reflective when turned to the light.

Flesh: cream; corky (dimitic); browning slowly on exposure to air.

Smell: resinous, like the resin used on Queensland surf boards.

Spore print: hyaline when fresh but cream if scraped together on a slide and dried.

Spores: fusiform; 6.08 – 8.47 × 2.33 – 4.12 µm, average 7.3 ± 0.8 × 3.1 ± 0.55, Q = 1.9 – 2.74, average Q = 2.38 ± 0.29; smooth, with globular oily contents.

Basidia: narrowly clavate; 15 – 20 × 4 – 5 µm, four spored.

Cheilocystidia: absent. There are sterile cells: Cunningham describes these as paraphyses.

Pleurocystidia: absent.

Pileipellis: a trichoderm.

Habitat: growing on a very large (> 180 cm diameter) hollow log; species could not be determined.

Notes: this species has normally been associated with a large underground sclerotium, but this collection was on wood. There are surprisingly few records for this fungus. The fungus was first described from near Toowoomba in Queensland in 1883.

Collections examined: PL180522, Triunia Reserve, Woombye, David Moss and Warwick Nash, 4 Jun 2022.

QUEENSLAND FUNGAL RECORD © Queensland Mycological Society

Original author: P. Leonard 12 June 2022

¹ Kornerup & Wanscher, *Methuen Book of Color* the standard for describing the colours of agarics.

Some Fungi from Cairns

Ray Palmer

Ray Palmer recently sent in a collection of exquisite photos, taken over several years, of fungi in the Cairns area. The notes are based on his emails. All photos © Ray Palmer – Ed.

***Mycorrhaphoides* sp.** The first group are of a *Mycorrhaphoides* sp. The first three images were photographed at Cheepi Creek, Cairns. I took

Matt Barrett to the spot where he collected specimens and identified them as such. The other image was photographed at my block back in 2010 with an old camera and shows the the small "teeth" underneath. This fungus is unusual in that it has a growth habit similar to *Podoserpula pusio*, forming a structure of multi-tiered brackets growing from a central stem.

A very similar species, *Mycorrhaphoides stalpersii*, has only recently been identified in India.



***Cuphophyllus* sp.** (Photos on the next page). On one of Matt Barrett's visits to my rainforest block west of Cairns to collect some fungi I mentioned to him that he might be interested in a "*Hygrocybe*" that had appeared close to my house. I first photographed this particular "*Hygrocybe*" on my block in 2010 and it has appeared at least three more times since.

Matt has confirmed that this "*Hygrocybe*" is in fact a *Cuphophyllus* sp. This is possibly new and a first record for Australia. The images were taken on days

2 and 13, and show how these fungi change over time. After approximately one week, the caps change colour from bright orange to yellow. The largest cap in the bottom left image measured 6.8 mm across. The seed is that of a foxtail palm. On day 14 the fungus was eaten by slugs. This species also occurs at Stoney Creek, some 4 km north of my block. One characteristic of *Cuphophyllus* is that the stipes are hollow. At this stage, I have found approximately 20 different *Hygrocybe* spp. in the Cairns region.



Other fungi

The following images are all from the Cairns area.

The following image of a large group of *Roridomyces irritans* (recently identified by Matt

Barrett) gives an idea of how prolific some fungi can be when conditions are right.



During early stages of growth, some trees in the rainforest are damaged and end up as two trunks growing from a single base. So it is with the fungi in the images below.

Top row: *Lentinus crinitus*, *Lentinus sajor-caju*,
 Second row: *Cyptotrama asprata*, *Ganoderma* sp.,
 Third row: *Mutinus* sp. (possibly *Mutinus bambusinus*), possibly *Delicatula integrella*.



At this stage I have not uploaded anything to iNaturalist, but I have uploaded well over 3,000 images of approximately 540 different species of fungi from the Cairns area on to the FABULOUS FUNGI-AUSTRALIA website. I am the moderator of this site which currently has 587 members. I am also

the main contributor to the site where I am known as "razor4343". Go to [flickr.com/groups/aussiefungi](https://www.flickr.com/groups/aussiefungi/).

All of my images are tagged and geotagged, and the vast majority can be enlarged to calendar or poster size without losing focus. Some are tagged as "unidentified" but I am gradually getting there.

How to Identify a Small Smelly Brown Fungus and Survive

Pat Leonard

Introduction

Many of us ignore small brown fungi: they have a well-deserved reputation for being difficult to identify in the field and being impossible to do anything with from a photograph. My general rule is to ignore them. One of the few exceptions I make is when they have a strong smell of garlic or rotten cabbage, because that takes you to a small group of a dozen or so species with a varied history that used to be in the genus *Micromphale*.

A bit of history

The genus *Marasmius* originated with Fries in 1836. Some 37 species were placed in this genus by Cooke (1892) in his review of the records of Australian fungi. Only three of those species might arguably be described as smelly. They were:

1. *Marasmius affixus*
2. *Marasmius foetidus*
3. *Marasmius putredinis*

The last two were reported from Queensland.

By the time that Cleland published his Larger Fungi of South Australia (1934/1935) he only confirmed *M. foetidus* from Cooke's list but added two further species:

4. *Marasmius australiensis*
5. *Marasmius cinnamomeus*

Singer (1951) moved *M. foetidus* to the genus *Micromphale*.

When Grgurinovic published her revision of Cleland in 1997 the genus *Micromphale* was still widely recognized. She transferred Cleland's *M. australiensis* and his *Cantharellus rugosus* to that genus and added two more species:

6. *Micromphale rugosum*
7. *Micromphale mirramirildinum*
8. *Micromphale villosipes*

Then, in 1997, Antonin and Noordeloos transferred some species out of *Micromphale* into *Marasmiellus* and this included *M. foetidus*. Halling et al transferred *M. villosipes* to *Gymnopus*.

Bruce Fuhrer's Field Guide to Australian Fungi (2005) has an image and short description of *Marasmiellus foetidus*.

Then in 2010, following the revision of the species concepts for Marasmioid and Collybioid fungi (Antonin, Halling and Noordeloos, 1997), *M. foetidus* was moved again, this time into *Gymnopus*.

In 2017, Petersen & Desjardin transferred *M. cinnamomeus* to the genus *Mycetinis*.

A winter foray

On the first day of winter in 2022 Judy Hewett collected a small brown fungus in the Maroochy Wetlands Sanctuary and, because it smelt of rancid boiled cabbage, I foolishly took it home as I thought it might be easy to identify.

Identifying a smelly small brown fungus

Since the advent of the Atlas of Living Australia it is easy (in spite of its omissions and errors) to get an idea of how common various species are, and whether they have been reported in Queensland. The table below records what I found for the eight smelly brown species listed above.

Observations of smelly marasmioid taxa in Australia, ALA, July 2022			
Species	Records in AU	Records in QLD	Last Qld record
<i>Marasmiellus affixus</i>	1467	3	2011
<i>Gymnopus foetidus</i>	16	1	2008
<i>Marasmius putredinis</i>	1	1	Circa 1880
<i>Micromphale australiensis</i>	0	0	1925
<i>Mycetinis cinnamomeus</i>	0	0	1924
<i>Micromphale rugosum</i>	0	0	1927
<i>Micromphale mirramirildinum</i>	1	1	2008
<i>Gymnopus villosipes</i>	7	0	2013

So, I was faced with a choice of eight species now spread amongst four genera. The next step was to try to get to a genus using Funkey. Using spore colour, habitat, substrate and odour one arrives at three possible genera: *Gymnopus*, *Heimiomyces* and *Leucocoprinus*. Activating the magic wand tells you that the next character that should be examined is the spores treated in Melzer's reagent. I do this and they are inamyloid, so we are in the genus *Gymnopus*.

An alternative key to the modern genera is given by Lassoë & Petersen's (2021) 'fungal wheel'. Adnate gills and strong smells get you down to *Gymnopus* or *Mycetinis*.

The Maroochy Wetlands collection had two distinctive features: it was caespitose, that is the stipes are joined together at the base, and it was growing in litter, not on a log.

Now we can dismiss *H. putredinis* which has not been seen for 140 years and has an inadequate description. *Marasmiellus affixus*, known as the little stinker, is a common small marasmioid fungus growing in troops on logs. It has an eccentric (off centre) stipe, so it can also be dismissed. We can also dismiss *G. villosipes* because it grows with Monterey pines, despite the fact that it was first described from Australia. That still left five options, three of which have not been seen since the 1920s.

Sadly, all five remaining potential species don't have information about growth habit and substrate.

Three of the remaining species are in Grgurinovic's helpful key to *Micromphale*, step 1 of which asks whether the cheilocystidia, that is the cystidia on the lower edge of the gill, are smooth or nodulose. A quick look at a cross section of the gill stained in Congo red reveals they are smooth and that rules out *M. rugosum*. A new description of *Mycetinis cinnamomeus* by Petersen and Desjardin in 2017 reveals that this also has nodulose cheilocystidia and can thus also be eliminated.

Grgurinovic's next couplet is based on spore size so I measure ten spores. Here are the results:

Species	Spore size	Q-value
<i>Gymnopus foetidus</i>	$8.7 \pm 1.8 \times 4.2 \pm 0.8$	2
<i>Micromphale australiensis</i>	$8.6 \pm 1.8 \times 4.2 \pm 0.5$	2
<i>Micromphale mirramirildinum</i>	$9.8 \pm 1.2 \times 6.4 \pm 1.4$	1.5
<i>Gymnopus</i> 'Maroochy wetlands'	$6.25 \pm 0.25 \times 3.05 \pm 0.26$	2.05

This analysis rules out *M. mirramirildinum*, which has significantly larger spores, cinnamon coloured lamellae and grows on *Eucalyptus* bark.

The most likely of the two remaining species is *Gymnopus foetidus*. There are 16 records for Australia, the latest from 2008, and only 2 of which are preserved specimens. But against this, one must consider the fact that this fungus was described from England, grows on wood not litter, has a strongly striate cap, and is associated with deciduous trees. *Gymnopus* 'Maroochy wetlands' is closer to *Gymnopus brassicolens* (of the European species) than it is to *G. foetidus*. We are fortunate to have

many images of *G. foetidus*, so we can also now make a visual comparison. Here are two images of *G. foetidus* from Europe:



Gymnopus foetidus © Mata & Petersen, Italy



Gymnopus foetidus © M. Noordeloos, England

Here are a selection of images labelled *Gymnopus foetidus* from the Atlas of Living Australia:



Gymnopus foetidus © P+B



Gymnopus foetidus © zeke 1944



Gymnopus foetidus © Reiner Richter

And here is the Maroochy wetlands collection:



Gymnopus sp. aff. *Micromphale australiensis*

My conclusion is that our collection is not *Gymnopus foetidus* and nor are most of the species illustrated on the Atlas of Living Australia with that name.

So, we are left with Cleland/Grgurinovic's *Micromphale australiensis*.

The identification process to this point is called 'shoehorning': trying to fit one's find to the closest published species description even though it is not an exact fit.

The shoehorning approach fails to recognise that there are a whole batch of undescribed species out

there. So, a further choice is whether you think that the find is:

1. Close enough to give it the remaining species name, in which case it is: *Micromphale australiensis*.
2. Near, but possibly a new species in which case it is: *Micromphale* aff. *australiensis* or (better) *Gymnopus* sp. aff. *M. australiensis*.
3. A new species in which case it gets a tag name: *Gymnopus* 'Maroochy Wetlands'.

Conclusions

If you find a smelly little brown mushroom I advise you to walk on by and save yourself a lot of time. If you want to try to find out what it is, be aware that the taxonomists have made more name changes in this area than you could poke a stick at. If you have the courage to proceed, record what you do so the next person doesn't have to go through all this again.

I decided to shoehorn this one purely because *Gymnopus* sp. aff. *Micromphale australiensis* seems a very sensible name for this smelly little brown mushroom lurking in a gloomy corner of the Maroochy Wetlands Sanctuary. Whichever name it is given, there will probably be a taxonomist lurking in a lab in the US of A or in Holland or the Czech Republic ready to correct it.

Note: Nigel Fechner notes that "At time of publication, the current name for *Marasmius putredinis* is *Hydropus putredinis*, and *Gymnopus villosipes* is *Collybiopsis villosipes* "

So be warned: This is never easy! (Ed)

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Litter Traps

The recycling lifestyle reaches new heights Frances Guard

Litter traps are collections of (mostly) dead leaves and twigs held together by fungal threads known as rhizomorphs. These threads are mycelia coated with melanin to create sturdy, water resistant, desiccation-resistant threads that can grow aerially between branches, twigs and leaves, entangling plants in the understorey and even in the canopy. The trapped leaves are then decomposed by the fungal rhizomorphs to provide it nutrients. This is very advantageous for the fungus as it avoids competition with the multiple cast of other litter rotters on the ground.

Rhizomorph-forming fungi are found across a number of genera, and are particularly well known in the *Marasmiaceae* family. This includes the horsehair fungus, *Marasmius crinisequi*, but it is not limited to that species or genus. I have recently become acquainted with another genus, *Crinipellis*, a relative of *Marasmius*, (in *Marasmiaceae*) forming huge litter traps in my rainforest on Dilkusha Nature Refuge. (See *Fungi of Queensland* for a full description) (<https://qldfungi.org.au/wp-content/uploads/2022/06/Crinipellis-sp.pdf>). By comparison, the litter traps of *M. crinisequi* are tiny, consisting of just a few dead leaves.

These *Crinipellis* traps are up to 0.6 cubic metres, and occur from 1.5 to 5 metres above ground. (Photos, right.) With all the recent rain they have been producing fruitbodies regularly along the twigs, and weighing the host branches down as a result. The host is *Cryptocarya sclerophylla*, a rainforest tree native to this area. The rhizomorphs travel inside twigs and on their surface and then become aerially oriented, attaching to adjacent living leaves. As dead leaves fall from the canopy they are trapped in this net and become fungal food. The heavy burden of fungus appears to be creating an unhealthy environment for the tree, which is dying back in places that have been



Crinipellis litter traps © Frances Guard.

sites for traps. It is not yet known whether the *Crinipellis* is actively killing leaves through production of volatile chemicals, or simply benefiting from dead leaf-drop. Continued observation may help to resolve this question. More photos on the next page.



Crinipellis upper and lower view of fruiting body. Note the rhizomorphs. © Frances Guard

An Invasive Fungus - *Favolaschia calocera*

Theresa Bint

The pretty little fungus below is *Favolaschia calocera* – also known as orange ping pong bats, orange conch or orange pore fungus. It is a weed fungus – an invasive species thought to have originated in Madagascar (though it has been suggested it was introduced there from Asia) and now found throughout the world.

A saprophytic (wood-rotting) fungus in the family Mycenaceae, it infests dead plant material and grows in clusters or troops. It's been recorded

growing on decaying hardwoods, conifers, bamboo and ferns as well as on untreated timber such as firewood and pallets. It is speculated that *F. calocera* arrived in Australia in imported timber.

Favolaschia calocera is a bright orange, fan-shaped fungus with a laterally attached stipe (stalk). The cap is 5 – 40 mm across, and has raised bumps which mirror the 'honey-comb like' pores on the under-surface. It is gregarious, meaning it appears in clusters or troops of up to hundreds of sporocarps (fruiting bodies). The spore print is white. The inclusion of the 5 cent piece in the image is a way of indicating scale. *Favolaschia calocera* can be widespread through a location – on tree trunks, twigs, fallen logs, as well as higher up in a forest canopy. Humans (and animals) are effective vectors for its spread: millions of airborne fungal spores attach to passers-by and distribute this invasive species. The spores settle on our hair, skin, clothes, boots, bags and equipment. Weedy fungi such as *F.*



Occurrence records map (454 records)



Occurrence of *Favolaschia calocera* in Australia

<https://bie.ala.org.au/species/4a70c0cb-148a-47ab-8ccf-7147a1a6dfba> Accessed 7/5/2022

calocera are a problem for our ecology as they may out-compete native species and become dominant, particularly in areas disturbed by human activity – along walking tracks and in national parks, for instance. (It seems that there have been no scientific studies to prove that as yet, but the danger cannot be ignored – Ed.)

It is such a photogenic species – bright colours ranging from orange to chrome yellow, crisp outline, honeycomb-like pores – many photographers of orange ping pong bats could have unwittingly carried its spores on their phones and cameras (and glasses, knives, and backpacks) to the next national park, reserve or area of bushland.

Some look-alikes in the fungi world

Fortunately, not all small and pretty orange fungi are *F. calocera*! Some possible look-alikes include, from left to right below, *Cyptotrama asprata* or golden-scuffy collybia (image, Theresa Bint), and two images of small gilled fungi (species unknown). A distinguishing feature of *F. calocera* is the honeycomb appearance of the pores under the cap.

What to do – Arrive Clean, Leave Clean!

You can help to limit the spread of *F. calocera*. Make sure your clothes, shoes and equipment are clean before you enter any bushland site, reserve or national park. Clean your gear afterwards – what can't be washed should be wiped or sprayed with methylated spirits or similar and/or put out in the sunlight. Infested plant material should be solarised (put in a black plastic bag and left out in the sun) for at least 6 weeks. Record your sightings of *F. calocera* in iNaturalist.

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All images by Catherine Mobbs, unless otherwise indicated. Locations: Mary Cairncross Scenic Reserve, Bunya Mountains National Park, and coastal paperbark swamp forest near Peregrin, South East Qld.

A new species name?

Zhang & Dai (2021) suggested that the species we have in Australia is now named *Favolaschia claudopus*. They describe specimens collected from the Dandenong Ranges in Victoria as being chrome yellow. Observations and collections in Mary Cairncross Scenic Reserve in Maleny and in the Bunya Mountains National Park in the last six months have been of very bright orange fungi – not yellow by any stretch. I suspect that we may have several species of this very invasive fungus and look forward to investigating this a bit further.

(I view this as speculative. I do not regard a simple colour variation as proof of anything other than a local variant. It is possible there was more than one introduction into Australia, but not enough sampling has been done to prove that one way or the other. The fungus I saw near Whanganui in New Zealand was bright orange - Ed.)

Reference

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