



THE QUEENSLAND MYCOLOGIST
Bulletin of
The Queensland Mycological Society Inc.

The Queensland Mycologist is issued quarterly. Members are invited to submit short articles or photos to the editor for publication. The deadline for contributions for the next issue is February 1, 2009.

Please ensure that the Secretary (secretary@qms.asn.au) always has your current email address. The Secretary, Queensland Mycological Society Inc, PO Box 295, Indooroopilly Qld 4068

SOCIETY OBJECTIVES

The objectives of the Queensland Mycological Society are to:

1. Provide a forum and a network for amateur and professional mycologists to share their common interest in macro-fungi;
2. Stimulate and support the study and research of Queensland macro-fungi through the collection, storage, analysis and dissemination of information about fungi through workshops and fungal forays;
3. Promote, at both the state and commonwealth levels, the identification of Queensland's macrofungal biodiversity through documentation and publication of its macro-fungi;
4. Promote an understanding and appreciation of the roles macro-fungal biodiversity plays in the health of Queensland ecosystems; and
5. Promote the conservation of indigenous macro-fungi and their relevant ecosystems.

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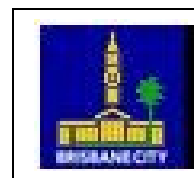
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QMS Website: www.qms.asn.au

Have you logged onto the QMS website lately? If not then it is time you did!! Many thanks to Andrew Kettle for getting the site up and running. Please provide feedback to the Committee about any ideas you may have for the site.

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QMS acknowledges and appreciates the sponsorship that has been given to the Society by the Queensland Herbarium, SEQ Catchments and Brisbane City Council.



QMS Calendar 2010

Members are reminded that 2010 subscriptions (\$20 pp) are due in January. Payment can be made by cheque mailed to the Treasurer, Queensland Mycological Society Inc., PO Box 295, Indooroopilly Qld 4068 or directly to the Treasurer at the next meeting. A Membership Renewal Form is included on page 17.

Meetings are held in the Bailey Room at the Herbarium, Mt Coot-tha, commencing at 7pm on the second Tuesday of alternate months from February, unless otherwise scheduled. Check the website for any changes.

There will be a pre-meeting at 6 pm. at the Herbarium at Mount Coot-tha for those who attended forays and took photographs. They are asked to bring all images preferably renamed according to the QMS naming convention summarised on the next page (see the website or March 20 Newsletter for details. The website will contain any updates to the protocol).

To assist those not in attendance at meetings, notes on the addresses given are included in issues of the Queensland Mycologist. However, the notes never do justice to the topic as they do not reflect the enthusiasm of the speaker or cover the questions and discussions that were raised on the topic. So remember, where possible it is far better to attend the meetings, get the information first hand and participate in the invaluable information sharing opportunity.

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QMS Meeting Programme 2010

Date	Speaker	Topic
9 February	Fran Guard	<i>Pleurotus tuber-regium</i>
13 April	David Fisher	Cultural attitudes to fungi. (to be confirmed)
8 June	John Wrench	Taxonomic Etymology
10 August	Andrew Kettle	Fungal Genetics
12 October	TBA	

Meeting Supper Roster for 2010

Two volunteers are required for each meeting – one to bring something savoury and one something sweet.

Meeting	Savouries	Cakes/sweets
Feb	Matthea Paulus	Fran Guard
Apr		Annita Hearle
AGM/May	All Bring a plate to share	
June	Andrew Kettle	
Aug	Pat Leonard	
Oct		
Dec	All Bring a plate to share.	

2010 Workshop Programme

Date	Topic	Leader
Saturday, 31 July		Patrick Leonard
Saturday, 28 August	Fungi photography for IDs	Sapphire McMullan-Fisher
Saturday, 25 September		Megan Prance

QMS 2010 Field Trip Programme

Date	Location	Leader
Saturday, 30 January	Linda Garrett Park, Mapleton	Sapphire McMullan-Fisher
Saturday, 27 February	Mount Cordeaux	Susan Nelles
Saturday, 27 March	Chermside Hills	John Wrench
Saturday, 10 April	Linda Garrett	Patrick Leonard
Saturday, 24-25 April	Springbrook A.R.C.	TBC
Saturday, 8 May	Samford Valley/ Redcliffe B.G.	Jan McNicol
Saturday, 29 May	TBA	Ray & Noreen Baxter
Saturday, 26 June	Southern Cooloola, Great Sandy National Park, Lake Cooroibah Road, QLD.	Sapphire McMullan-Fisher
Saturday, 30 October	Northern SE Qld	Patrick Leonard
Saturday, 27 November	Mushroom Farming	Jon Atkinson / Kim Nguyen

QMS has a system for consistent naming of images. Go to the members section of the website for information. While the system looks complex, it allows for quite a lot of information to be saved in a computer-sortable form, and that makes it a lot easier to collate images later on.

Long Term Foray Programme:

There are five long-term regular foray study sites covering the ecological diversity that reflects the main habitat types in South East Queensland. A mixture of day and weekend forays are run. These sites rotate dates so that they are visited in each month of the season. The five long term regular sites are:

Sites	Habitat	Leader	Visits
Springbrook ARC	regeneration	Diana Leemon	24-25 April, 10; 28 - 30 Nov., 08; 2 - 4 May, 08
Linda Garrett Park, Mapleton	wet sclerophyl	Patrick Leonard	30 Jan, 10; 5 April, 08; 10 Nov., 07
Chermside Hills	dry sclerophyl	John Wrench	27 Mar, 10; 25th April, 09 5 Jan., 08; 7 July, 07

Cunningham's Gap	Rainforest	Susan Nelles	25 October, 08 6 October, 07
Great Sandy National Park	wallum/sand dune	Sapphire McMullan-Fisher	26 June, 10

President's Report

Sapphire McMullan-Fisher

Best wishes of the season to you all. For those of you who couldn't make it we had a wonderful end of year party on Tuesday 8th Dec 2009. Sadly there were less entrants this year so we didn't take donations for the library. None the less -this years great winners were:
 Best Image Prettiest = Ruth Thomson
 Best Ugly Fungus = Fran Guard
 Best Spoken Word/Song = John Wrench
 Best Limerick = John Wrench
 Best fungi food – savoury = David Holdom
 Best fungi food – sweet = Rhoda Warhurst

We all enjoyed some of Taylor Lockwood's art. For those who don't know Taylor he is a great photographer and musician who has been bitten hard by the fungal bug. Check out his weird and wonderful website about fungi at <http://www.fungiphoto.com/> and <http://www.kingdomoffungi.com/>

Before the really festive part of the meeting got started Fran and Sapphire gave an update on the Fungi Museum Book. To find out more read the article 'Fungi needed for Museum book' (in this Newsletter) this include a list of priority species. In a nutshell we are hoping that members will spend the following fungi season getting high quality images with collections of fungal species. Some advice about getting useful fungi images please read '*Some ideas on Fungal Photography*' (in this Newsletter). To help members make good collections to be voucher specimens of their images please read and refer to the 'Fungi Collection Manual' in the members section of the QMS website (will be up by January). Patrick put together the manual and I wish I'd had such a comprehensive guide when I was getting started with the fungi. If members would like a printed copy of the manual posted to them they need to email me with their postal address or pick them up at the next meeting. I expect the print costs will be about \$5 and postage \$2.50. For those of you who also have got copies of the Largent books are also great reference material. If members need advice about collecting they should contact Sapphire president@qms.asn.au or Patrick vicepresident@qms.asn.au.

As most of you know the last two forays to Springbrook and Linda Garret had to be cancelled due to the dry conditions. We had about 45 mm two weeks ago and a few puffballs and lawn Agarics popped up but I haven't seen any bush fungi yet. Let's hope we get more rain or it will be a poor summer fungal season.

If people are in doubt about whether an activity is happening you should first check the website <http://www.qms.asn.au/>. For forays if you have signed up the foray leader will contact you if it is cancelled, you can also contact Andrew the Foray Coordinator if you cannot contact the foray leader.

That's all for now. I hope to see you at the January foray or at the next meeting on February 9th 2010. Until then best wishes for a happy and safe festive season and new year.

Some ideas on Fungal Photography

Sapphire McMullan-Fisher

Let me start by saying I am not a 'professional' photographer – so the ideas and suggestions that follow are simply stuff I have picked up or worked out in the 14 years I have been focused on photographing fungi. In that time the technology has changed dramatically. I started in film, moved to slides and now work entirely in digital! So for those who still work in slide or film - I'm afraid most of my comments are about digital photography.

Some basic tips:

Use macro settings – all but the biggest fungi will look better if macro settings are used. If you can use an 'aperture priority' setting as this allows you to take images with a greater depth of field – i.e. let you get the stem and cap both in focus. Use the highest F-stop you can which slows the image down, so most shot's cannot be hand held hence the next point. The best combination is the macro setting in aperture priority mode.

Use a timer and tripod – most fungi images are taken in lower light conditions so use of a tripod and timer will let you take images without your camera shaking. Even if you are not using a tripod, use of the timer means you don't get the blur from the camera shake when you press the button. There are a number on miniature tripods, stands, and bean bags on the market.

Keep out of the sun – although you want as much light as you can, sunlight can cause too much contrast, so shade your subject – I use my hat or get someone to stand in the direct sunlight. Overcast days are the best for fungal photography – the fungi don't shrivel as fast!

Take care with flashes and lights – some fungi particularly shiny ones can reflect light. Direction flash can give you odd bright spots and shadows. I recommend the use of separate lights (flashes) if you are working in low light conditions. Always take a couple of shots without the lights to see what the natural colours of the fungus were.

Take your time – I know on foray's it can be hard with everyone wanting to get a picture of all the fungi but if you can – try to take your time. Let others take a quick snap and move on – just make sure they don't mess up the fungus!

Take as many shots as you need – digital cameras are great if you think you messed up take another image – remember to delete poor and average shots when you download otherwise you'll chew up memory space.

Use high resolution setting – you can always make a smaller copy of images to send on email or use on the web or whatever but you can never make an image bigger (higher resolution). I keep all my high resolution images in a folder by date and make lower resolution copies to use in everyday things. If I mess up an image or it's too small for what I need I can always go back to the original.

Get organised and be consistent – there is nothing more frustrating than knowing you have an image of something but not being able to find it. I recommend developing a system and sticking with it. Using programs which help you organise you images can be a great help but you must be consistent with the terms you use and keep notes on dates, places and species you have seen. For QMS stuff **always** take an image where the tag number is readable. With file naming the QMS system really does help you and others keep track of when and where an image was taken and this allows you to find out the best name for the fungus (more on this later). Don't forget to have a regular, external **back up system** – you'll probably never see some of these fungi species ever again!

Different types of fungi shots



Figure 1. GPS screen with location



Figure 2. Laboratory style shot of *Dermocybe splendida* on white paper with ruler.

There are different types of images which are used/needed/wanted for different reasons. What follows is a system which allows you to take different types of shots, if you are serious about naming fungi you will need to take what I call fungus ID (identification) shots.

Take an image of the tag or number of the fungus, this can be a close up just of the tag or piece of paper. Some people also write down the date and place, others then take a photo of their GPS screen (Figure 1) so they also have a copy of the geographic coordinates. You will need to track the information about the fungi you are taking. Note the fungus is not in these first few shots.

Aesthetic/Natural images – these images are trying capture the natural beauty of the fungus and it's surrounds. Try not to mess about with the fungus too much at this stage – take a few shots from different angles before you start rearranging the leaf litter, grass etc. Then you can move unsightly blades of grass etc, you may also want to try something more artistic. This is also a good time to take **habitat** shots of the setting and substrate before you disturb the scene too much.

ID shots 'natural' – these are images which help identify the characters of the fungus but still look relatively natural. A little 'gardening' i.e. moving the fungus around so that you can see the top and bottom surfaces in the same shot. Sometimes you might even cut one of the fungi in half to see the internal characters but this getting into the next category. Some people use a small paint brush to tidy up their fungi – take care not to damage your specimens too much.

ID shots 'taxonomic' – in these images the priority is getting as much information about the fungus in the one or two shots. Try to get all developmental stages in the view, this is often the point where you cut the fungus in half or touch it to show that it bruises etc. You should also include the tag (fungus field number), and a scale like a ruler or a coin (Figure 2). A white item can be very useful for at least one of the images as a 'white point' can be important for colour and light correction.

ID shots 'laboratory style' – these shots may be taken at the site, in the car park or at home. If you are taking the images at home I still suggest using as much natural light as possible. For the background try to use white or ideally 25% grey. Wooden or coloured table-tops can be very distracting. Again the aim here is to get the different fungal stages and

characters in the shot – this time under more consistent conditions. If I can't use daylight I use daylight lamps. If using lamps be careful they don't make big shadows.

Image sizes, file types, resolutions etc.

As I said before you can't make images bigger so it is best to take images in the highest resolution and of the largest size you can. If you have a higher end camera it may be best to take your shots in raw (not all cameras will do that) and then process them for other uses - the software and instructions should come with your camera.

Images for computers

The resolution of computer screens varies, depending on the screen and individual settings. Newer screens have higher resolutions, but some people do not like the small icons that result and set them to lower resolutions. That suits smaller files sizes which is good if you want to email your images or share them on the web as it takes less time to upload them. However low resolution images will look pixelated if you try to zoom in. Most screens are now at least 800 x 600 pixels and most are now more like 1280 x 1024 pixels. Other typical resolutions are 1650 x 1050 and 1440 x 900.

Images for Print

When images are used for print media they need to be at a minimum resolution of 300 dpi and size of 13 cm (equivalent to about 1500 pixels). For books and the like we may want to zoom in and crop just part of the image. For this reason we ask that you give us images with the largest resolution and size possible. For example we would prefer 600 dpi and 50 cm (about 12,000 pixels) even if it is 12 megabytes (MB) over the same image at our minimum standard (300 dpi and 13 cm).

A warning about JPEGs

If you open JPEG images just to view them do not save them as part of the closing process. If you do, this will progressively reduce the quality and file size. JPEG images are compressed, but that loses some detail. Resizing loses even more detail. An uncompressed image is a bitmap, but a 10 megapixel bitmap in 32 bit colour would be 40MB! GIF images are compressed, but without data loss, so are intermediate in size, but are less common these days.

Naming images

Images given to QMS for the book and other purposes need to be renamed using the QMS image naming system (the current version of instructions are on the web). If the image is not from a QMS foray please use a 'P' instead of a 'Q' after the date number. We will also need species, location and collection details (for collections refer to the collection manual). If the images are taken on a QMS foray we just need the correctly named files as these will tell us the date and hence the location, the field number will give us the species and your initials in the file name will tell us whose image it is!

Most people should be able to rename their image files as part of the process. When you download your photographs to your computer, you may be given an option to name them. Just put in the number format for each day's download then later you just have to edit the field number for each image e.g. 2091124Q_SMFF – would be all the images of mine taken on a QMS foray on 24 Nov 2009. In Nov 2010, the format will be 2101124Q. On some forays we may find over 100 specimens, so the Q part of the name will be Q001 for specimen 1, Q010 for specimen 10. Check the field tags in the images to see how to fill in the Q number. This will allow you to check the name of the fungi with the file we put up on the QMS website. It will also allow us to put your photo in the database and match it with the correct information.

Foray Report-Redcliffe Botanic Gardens July 2009

The Redcliffe Botanic Garden is a former dairy farm that was converted to a botanic garden only 30 years ago and is run by volunteers. It contains a wide range of native trees and it is run to be as natural as possible. Fallen wood is left *in situ*, so it is a good site for wood rotting fungi.

Few agarics were seen- this *Coprinus* (fig 1) is in the *plicatilis* group and is probably here due to high nitrogen from bat guano from the fruit bats colony that roosts in the gardens.



Figure 1: *Coprinus* sp.

Pycnoporus coccineus (Fig 2) was the most common polypore on dead wood, and it was also found on living trees (Fig 3).



Figure 2: *Pycnoporus coccineus* on dead (left) and living (right) wood

Another fungus was a fleshy-gilled *Laccaria* growing in the sand near an *Acacia*, and identified only as "species E" (Fig 3).



Figure 3. *Laccaria* "sp. E". Note two spored basidia in bottom photo.

Other notable finds were *Microporus xanthopus* on a branch of *Mallotus discolor* (Fig 4) and *Phellinus badius* on a log (Fig 5) as well as an unidentified *Coprinus* (on wood), *Cyathus striatus* (on wood) *Ganoderma australe* (on the trunk of an *Acacia*), *Hexagonia tenuis* on the branch of an unidentified tree, *Pycnoporus coccineus* and *Trametes lactinea* (on *Eucalyptus* branches), *Schizophillum commune* and *Steccerinum* sp (both on twigs), and *Trametes hirsuta* and *Trametes versicolor* (on logs).



Figure 2: *Microporus xanthopus*



Figure 3: *Phellinus badius*

How To Become A Serious Amateur Mycologist In Five Easy Steps

Pat Leonard - August 2009

Step 1 Adopt a genus.

This step is the most difficult, so choose very carefully. You have two options:

- a large genus which is well documented like *Mycena* or *Amanita* but which has not been studied in Queensland.
- a small genus with at least 4 and less than 10 species which no one else is interested in. Small brown mushrooms that other people stamp on can often yield high dividends.

Choose something that is attractive to you and has at least one or two common species and get to know these. You can then impress other forayers by learning to spot the other species in the genus. You will be surprised how easy this becomes with a little practice. Do not try to start with a large badly documented genus like *Cortinarius*, you will become depressed and may need to seek medical attention.

Step 2 Get a description of the genus

For gilled basidiomycetes you can usually find a description in:

- Knudsen & Vesterholt (2008) *Funga Nordica*. (In QMS Library)
- Largent & Baroni. (1988) *How to identify mushrooms to Genus Vol VI The modern genera*. (In QMS Library)
- Singer (1986) *The Agaricales in Modern Taxonomy*
- Grgurinovic, C. (1997) *Larger Fungi of South Australia*.

For the polypores you will find a description in:

- Ryvarden, L. (1991) *Genera of the Polypores*.

For the gasteromycetes you will find a description in:

- Miller, O.K. and H.H. (1988) *Gasteromycetes*. (In QMS Library)
- Cunningham, G. (1942) *Gasteromycetes of Australia and New Zealand*. (In QMS Library)

Get one or more descriptions of the characters of your chosen genus. Ask Pat or Susan to help you get these. When you have them, open a file on your computer with the name of your chosen genus, and a matching paper file. Make sure you memorise the key characters of your chosen genus. Normally this will include the general shape and size of the fungus, its spore colour and habitat.

Step 3 Find out what species have been reported in Australia.

This is the easiest step of all and should not take you more than 1 hour. Borrow:

- *Fungi of Australia Volume 2A* for most basidiomycetes (In QMS Library)
- *Fungi of Australia Volume 2B* for polypores and gasteromycetes. (In QMS Library)

You can obtain the same information from the Interactive Catalogue of Australian Fungi on the net. Go to:

- http://www.rbg.vic.gov.au/research_and_conservation/fungi/cat/search and type in the name of your genus and then place a % in the species name. For example:

Genus = Copelandia
Species = %

You will soon discover that there may be several names that have been used for a single species (synonyms). The beauty of the interactive catalogue is that it sorts this out for you and gives you the current name (**in black**) and the synonyms (**in red**). Make a copy of the list including the synonyms and place the list in each of your files.

Step 4 Find out which species have been reported in Queensland

You do this in two stages, stage 1 is easy. Go to the Australian Virtual Herbarium (AVH) website at:

- <http://www.rbg.vic.gov.au/avh/>

Hit the data access button. The 'data search interface' screen comes up and in the 'genus' box enter the name of your genus and in the species box enter %. In the 'Search the following herbaria' box uncheck all the herbaria except Queensland so that you only have a green tick mark in the Queensland box. Hit the 'search records' button and a 'data formatting interface' screen will appear telling you how many records have been found. Now hit 'submit query'. A screen will appear with a list of all the collections in your chosen genus that are in the Queensland Herbarium at Mt Coot-tha together with their reference numbers. Some of these may be old names, but you will know the correct names from the synonyms information you got in step 3. Make a copy of the list and place in each of your files.

Stage 2 involves going in to the Herbarium with Pat or Sapphire and going through the collections, but I strongly recommend you leave this step until you have completed your file and made some of your own collections.

Step 5 Get full descriptions and images for each species.

You now have a list of the species in your chosen genus that you expect to find in Queensland! The next step is to get a full description and an image for each species. The place to start for gilled basidiomycetes is:

- Grgurinovic, C. (1997) Larger Fungi of South Australia.

For the polypores you may find a description in:

- Cunningham, G. (1965) Polyporaceae of New Zealand (in QMS library).

For the gasteromycetes you may find a description in:

- Grgurinovic, C. (1997) Larger Fungi of South Australia.
- Cunningham, G. (1942) Gasteromycetes of Australia and New Zealand. (In QMS library)

There may also be images and short descriptions in:

- Fuhrer, B. (2005) A Field guide to Australian Fungi. (in QMS library).
- Grey, P and E. (2001) Fungi down under. (in QMS library).
- Bougher, N. and Syme, K. (1998) Fungi of Southern Australia. (in QMS library).

You can also try for images by:

- asking Megan if there are any images in the QMS image library.
- doing a Google image search for your species in 'pages from Australia'. But be warned, you will get many incorrectly named images and many non Australian ones.

If at Step 1 you chose to work on a large well documented genus, you will have to acquire the relevant literature. Susan and Pat can advise you on this.

What to do next

It is worthwhile spending a little time (a couple of hours on a wet day) getting your file in to some sort of order so that ideally you end up with one page with the description of the genus and a key to the species if you have found one, and then a separate page for each species which should look a bit like the Fungi of Queensland pages with a photo or painting at the top followed by a description of the field characters and then the microscopic characters.

Now you are ready to go out in the field and hunt for your fungi. When you find them:

- make a good collection, 4 or 5 fruiting bodies and dry them
- take photographs that show the main characters
- make a spore print and
- write a full description.

After a year or two you may need to think about revising your original descriptions that you got from books or the net. This is the point at which you need to go to the Herbarium and check the material there (Step 4 stage 2).

If you get here you will probably now know more about your chosen genus than anyone else in Queensland!

Fungi Needed for Museum Book

Fran Guard

The basic goal of this little book will be to describe and illustrate some of the common, as well as some of the more interesting, uncommon fungi of SE Qld. It will be aimed at the interested but ignorant public, as an introductory educational tool.

Although we have some images and collections from forays for a publication we need high quality images. To facilitate identification and verification of each species illustrated we need collections of the fungi in the book to be deposited in the Brisbane Herbarium. We need high quality images for the book (see article on how to take fungi images in this issue). The upshot of this is that photographers need to do proper collections for the species we want in the book. Members should refer to the Collection Manual for instructions on how to do a proper collection. If you need advice about making collections please contact Sapphire (email: president@qms.asn.au or 5485 3066) or Patrick (email: vicepresident@qms.asn.au).

The table below (Table 1) is a list of the fungi we would like images and collections of. The table is ordered in priority and by morphological groups*. We have supplied page numbers from the common field guides (FDU = Fungi Down Under, Y= A.M. Young 2005, F = B. Fuhrer 2005, FOW = Fungi Out West). We also have reference material for some of these fungi in Fungi of Queensland (FoQ). If you would like the FoQ information on a particular species please contact Fran (email: guardphilpot1@bigpond.com). We also hope to put these up in the members section of the QMS website. This table could be printed out and carried in the field so you can see if a specimen is on our priority list.

References:

Chinchilla Field Naturalists Club (2007) 'Fungi Out West: some fungi of southern inland Queensland.' (Chinchilla Field Naturalists Club: Chinchilla, Qld)

Fuhrer B (2005) 'A field guide to Australian Fungi.' (Bloomings Books: Melbourne)

Grey P, Grey E (2005) 'Fungi down under.' (Fungimap: Melbourne)

Young AM (2005) 'A Field Guide to the Fungi of Australia.' (University of New South Wales Press: Sydney)

Table 1. List of macofungi for Museum Book. Quality images and collections are needed

*	Name	FoQ	Page Nos
Birdsnest			
Z	<i>Cyathus</i> sp	No	Y05-205, F05-222, FOW-137
Ascomycetes			
X	<i>Daldinia concentrica</i>	yes	F05-315
X	<i>Xylaria grammica</i>	yes	
X	<i>Xylaria polymorpha</i>	No	Y05-228, F05-342, FOW-28
A	<i>Chlorociboria</i> sp.	No	FOW-20
A	<i>Cyttaria septentrionalis</i>	No	FDU-103, Y05-222, F05-314
A	<i>Microglossum viride</i>	No	Y05-223, F05-327
A	<i>Plectania campylospora</i>	No	FDU-109, Y05-225, F05-335
A	<i>Scutellinia</i> sp	No	F05-337
Corals			
R	<i>Clavaria miniata</i>	No	Y05-86, F05-198, FOW-97
R	<i>Ramaria lorithamnus</i>	No	Y05-91, F05-211
R	<i>Deflexula fascicularis</i>	No	
R	<i>Hericium coralloides</i>	No	FDU-75, Y05-89, F05-241
Polypores			
P	<i>Amauroderma rude</i>	yes	
P	<i>Dictyopanus pusillus</i>	No	FDU-64, Y05-71, F05-153
P	<i>Favolaschia calocera</i>	yes	FM website
P	<i>Filoboletus manipularis</i>	No	Y05-134, F05-69, FOW-52
P	<i>Fomitopsis lilacinogilva</i>	No	Y05-73, F05-252, FOW-126
P	<i>Ganoderma australe</i>	yes	Y05-73, F05-253, FOW-113
P	<i>Grifola colensoi</i>	yes	F05-256
P	<i>Hexagonia tenuis</i>	No	F05-259, FOW-118
P	<i>Laetiporus sulphureus</i>	Find	Y05-75, FOW-121
P	<i>Lenzites acuta</i>	yes	
P	<i>Microporus affinis</i>	No	FDU-71, F05-265
P	<i>Microporus xanthopus</i>	No	FDU-70, Y05-76, F05-266, FOW-122
P	<i>Polyporus arcularius</i>	yes	Y05-79, F05-268, FOW-124
P	<i>Pycnoporus coccineus</i>	No	Y05-80, F05-271, FOW-125
P	<i>Ryvardenia campyla</i>	yes	F05-272
O	<i>Trametes versicolor</i>	No	Y05-81, F05-274, FOW-126
Leathers			
O	<i>Cymatoderma elegans</i>	No	FDU-77, Y05-82, F05-279, FOW-107
O	<i>Podoscypha involuta</i>	No	FOW-108
O	<i>Stereum ostrea</i>	Yes	FDU-79, Y05-83, F05-285, FOW-111

	Name	FoQ	Page Nos
Toothed			
J	<i>Hydnum repandum</i>	yes	Y05-94, F05-242
Jelly fungi			
N	<i>Auricularia auricula-judae</i>	yes	Y05-6, F05-289, FOW-100
N	<i>Auricularia cornea</i>	yes	Y05-64, F05-290, FOW-101
N	<i>Auricularia delicata</i>	yes	FOW-102
N	<i>Calocera</i> sp	No	Y05-65, F05-290, FOW-103
Stinkhorns			
M	<i>Aseroe rubra</i>	No	FDU-95, Y05-193, F05-228, FOW-138
M	<i>Phallus multicolor</i>	No	Y05-201, FOW-141
M	<i>Phallus rubicundus</i>	No	Y05-202, F05-235, FOW-141
Puffballs			
L	<i>Calostoma fuscum</i>	yes	FDU-90, Y05-203, F05-225
L	<i>Geastrum saccatum</i>	yes	
L	<i>Geastrum triplex</i>	no	Y05-208, F05-217, FOW-132
K	<i>Morganella purpurascens</i>	yes	
K	<i>Morganella pyriforme</i>	No	Y05-210, F05-214, FOW-134 as Lycoperdon
K	<i>Pizolithus</i> sp	yes	Y05-212, F05-219, FOW-145, FOW-146
Boletes			
H	<i>Austroboletus lacunosus</i>	yes	F05-187
H	<i>Boletellus emodensis</i>	yes	Y05-187, F05-188
H	<i>Boletus</i> sp 16	yes	
H	<i>Gyroporus caespitosus</i>	yes	
H	<i>Phlebopus marginatus</i>	No	Y05-188, F05-192, FOW-93
H	<i>Phlebopus xanthopus</i>	yes	
H	<i>Phylloporus hyperion</i>	yes	
H	<i>Tylopilus queenslandianus</i>	yes	
Gilled			
G	<i>Agaricus austrovinaceous</i>	yes	F05-18
G	<i>Agaricus campestris</i>	yes	Y05-101, FOW-33
G	<i>Agaricus xanthodermus</i>	no	FDU-17, Y05-101, F05-18, FOW-34
G	<i>Amanita albobolvata</i>	yes	
G	<i>Amanita flavella</i>	yes	
G	<i>Amanita ochrophylla</i>	no	Y05-107, F05-25
G	<i>Amanita punctata</i>	yes	
G	<i>Amanita pyramidifera</i>	yes	
G	<i>Anthracophyllum archeri</i>	yes	FDU-22, Y05-111, F05-28, FOW-38
G	<i>Armillaria luteobubalina</i>	no	FDU-23, Y05-112, F05-29, FOW-39

	Name	FoQ	Page Nos
Gilled			
G	<i>Bolbitius vitellinus</i>	yes	FDU-25, Y05-114, F05-32, FOW-40
G	<i>Cantharellus concinnus</i>	yes	Y05-95, F05-35
G	<i>Chlorophyllum molybdites</i>	No	Y05-117, FOW-42
G	<i>Clitocybe australiana</i>	yes	
G	<i>Collybia eucalyptorum</i>	yes	F05-38
G	<i>Conocybe lactea</i>	No	Y05-119
G	<i>Coprinellus disseminatus</i>	No	Y05-122, F05-41, FOW-45
G	<i>Coprinus atramentarius</i>	No	Y05-120, F05-43, FOW-44
G	<i>Coprinus truncorum</i>	No	Y05-123, F05-41
G	<i>Cortinarius archeri</i>	No	F05-44, Y05-124, FOW-46
G	<i>Cortinarius rotundisporus</i>	No	FDU-30, F05-48
G	<i>Gymnopilus junonius</i>	yes	FDU-37, Y05-135, F05-97
G	<i>Humidicutis arcohastata</i>	yes	
G	<i>Humidicutis lewellinae</i>	yes	FDU-41, Y05-140, F05-87
G	<i>Hygrocybe chromolimonea</i>	No	
G	<i>Hypholoma brunneum</i>	No	F05-96
G	<i>Hypholoma fasciculare</i>	No	Y05-143, F05-96
G	<i>Inocybe asterospora</i>	yes	
G	<i>Inocybe australiensis</i>	yes	F05-99
G	<i>Inocybe griseolilacina</i>	yes	
G	<i>Laccaria lateritia</i>	no	Y05-146, F05-102
G	<i>Lactarius aff. glaucescens</i>	yes	
G	<i>Lactarius clarkeae</i>	yes	Y05-147, F05-103
G	<i>Lactarius eucalypti</i>	yes	F05-104
G	<i>Lentinula lateritia</i>	yes	FOW-58
G	<i>Lentinus sajor-caju</i>	yes	
G	<i>Lepista sublilacina</i>	yes	Y05-151
G	<i>Leratiomyces ceres</i>	No	F05-95 as <i>Hypholoma aurantica</i>
G	<i>Leucocoprinus birnbaumii</i>	No	Y05-152, F05-114, FOW-61
G	<i>Leucocoprinus cepaestipes</i>	yes	
G	<i>Macrolepiota dolichaula</i>	yes	Y05-154, F05-118, FOW-66
G	<i>Marasmius haematocephalus</i>	No	
G	<i>Melanoleuca polioleuca</i>	yes	
G	<i>Mycena leiana var australis</i>	yes	FDU-48, Y05-159, F05-135, FOW-71
G	<i>Mycena sp luminous</i>	No	
G	<i>Mycena toyerlaticola</i>	yes	F05-142
G	<i>Mycena viscidocruenta</i>	yes	FDU-50, Y05-161, F05-143, FOW-72

	Name	FoQ	Page Nos
Gilled			
G	<i>Omphalotus nidiformis</i>	No	FDU-53, Y05-162, F05-182, FOW-92
G	<i>Oudemansiella canarii</i>	yes	Y05-163
G	<i>Panus fasciatus</i>	No	FDU-55, Y05-168, F05-155, FOW-87
G	<i>Pleurotus tuber-regium</i>	yes	FOW-88
G	<i>Russula erumpens</i>	yes	Y05-176
G	<i>Russula neerimea</i>	yes	
G	<i>Russula</i> sp	No	
G	<i>Russula viridis</i>	yes	
G	<i>Schizophyllum commune</i>	No	FDU-57, Y05-179, F05-172, FOW-89
G	<i>Stropharia semiglobata</i>	No	Y05-181, F05-174
G	<i>Tetrapyrgos nigripes</i>	yes	
G	<i>Tricholoma eucalypticum</i>	yes	Y05-182, F05-176
G	<i>Tricholomopsis</i> sp	yes	On bunya wood similar to Y05-183, F05-177, FOW-83
G	<i>Trogia umbrinoalba</i>	yes	Similar to F05-178
G	<i>Volvariella pusilla</i> var. <i>Taylori</i>	yes	
G	<i>Xerula trichifera</i>	yes	Similar to FDU-54, Y05-186, F05-180

Ref: FDU = Fungi Down Under, Y05 = Young 2005, F05 = Fuhrer 2005, FOW = Fungi Out West, page no after hyphen.

* Morphological group code: A= Asco, X=Ascospores, G=Gilled, H=Bolete, J=Toothed, K=Puffball, L=Puffball, M=Stinkhorn, N=Jelly, O=Leather, P=Polypore, R=Coral, and Z=Birdsnest.

Queensland Mycological Society Inc

ABN 18 351 995 423

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Email Address:.....

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